President’s Corner

Bird Flu... I know, I know, it’s not the correct name to use, but it is what most people know and ask us about. By now just about everybody in the wildlife disease community is dealing with avian influenza (AI) in some form or another. While there currently is not a human pandemic, there is global concern about what might happen over the coming months. However, from the poultry point of view, highly pathogenic avian influenza H5N1 is really serious. Millions of poultry have either died from virus infection or from culling operations designed to stamp out the virus. As wildlife disease folks around the globe, we are being barraged with questions about AI. What is the role of wild birds in the spread of this disease? When will the virus arrive in our area? What does the virus look like in wild birds? How do I protect myself from getting the virus if I hunt, watch, feed, eat, or work with wild birds? And so on. I certainly do not wish to make light of the situation because in some parts of the world, highly pathogenic avian influenza H5N1 is serious business with huge losses in poultry and losses in human life. Indeed, poultry is a primary source of protein in many parts of the world where highly pathogenic avian influenza H5N1 is active and so the direct loss of this source of food has had dramatic economic and nutritional consequences.

So far, AI (that is, highly pathogenic avian influenza H5N1) has not been reported in the Western Hemisphere. In the rest of the world, the virus has moved quickly across several continents and infected both wild and domestic birds. No one is really sure how the virus moved as quickly as it did. There is speculation and some data that suggest virus movement was facilitated by commercial activities associated with the poultry industry. Others suggest the virus moved with infected migratory waterfowl. Interestingly, there have been numerous reports of the highly pathogenic avian influenza H5N1 killing wild birds. The numbers of dead birds and species have varied, but for the most part these die-offs involved waterfowl or shorebirds. It is unusual for avian influenza viruses to kill wild birds. A number of die-off events have involved relatively few birds. This has led to some considerable consternation in the United States.

Several of the Western Hemisphere nations have launched surveillance programs to detect the virus as early as possible within a geographic area. These efforts vary considerably and some have questioned their utility. Government sponsored efforts in the United States have focused on five strategies to detect the highly pathogenic avian influenza virus H5N1. They are: surveillance of live-captured wild birds and hunter-harvested wild birds (including subsistence hunters), investigation of wild bird die-offs, testing of environmental samples (basically bird poop), and monitoring of sentinel flocks of birds. Understandably, the portion of the world free of the virus is watching the rest of the world that has the virus. So reports of highly pathogenic avian influenza H5N1 in one or two dead wild birds in Europe has folks tasked with looking for dead wild birds in the United States wondering if every dead bird they find is “the one”.

The Wildlife Disease Association does not regard the Supplement to the Journal of Wildlife Diseases (Wildlife Diseases Newsletter) as a citable publication and, therefore, it should not be referenced in the scientific literature.
Once again, around the world, wildlife disease folks are struggling with providing a perspective on the importance of wild birds, the commonality of avian influenza viruses in wild birds (especially waterfowl and shore birds), and how to best help others tasked with protecting poultry and people. Wild birds are not a threat to the stability of global society and culling wild birds to prevent this disease is not an option.

There is a bright side to all of this. First, there is the real possibility that the highly pathogenic avian influenza H5N1 will not readily convert to people (there seems to be some growing evidence of this) and a pandemic from this virus will not occur. Second, by conducting all these tests of wild birds, scientists will have an increased understanding of the prevalence of avian influenza viruses in wild bird populations, they will gain further understanding of bird movements, interactions between migratory birds and residents (mixing), persistence of the virus in the environment (or not), interactions between wild birds and poultry, and much more. This information will add to the large body of information already available on avian influenza viruses as a result of the work of several scientists (some are members of the WDA) around the world.

Will there be a pandemic? Hard to say. Many very smart people around the world say it is not question of if but when. If there is a pandemic, will it be the result of transformation of the highly pathogenic avian influenza virus H5N1 into humans? Hard to say that either. There are at least two other avian influenza viruses that appear to be moving into people now, so they could be candidates for the next pandemic as well. Regardless, wildlife disease professionals will remain at the forefront of this global disease.

-Scott D. Wright, President

WDA NEWS

The 55th Annual Meeting of the Wildlife Disease Association. The 2006 Annual WDA Conference at the University of Connecticut this past August was one that will be remembered for many things, but it will especially be remembered for celebrating new faces. There was nearly a record attendance (one shy!) and it included many folks who had never before been to a WDA meeting. And when asked if they would come back, the universal answer was a resounding “YES!” This year’s theme “Advancing Global Health: Facing Disease Issues at the Wildlife, Human, and Livestock Interface” brought in many excellent papers and posters. WDA’ers had the pleasure of hearing Dr. Gary Wobeser speak about the evolution of wildlife disease study as the Carlton Herman Founders Fund Lecturer. Other outstanding moments of the conference included the Auction, during which $6638 was raised, over half of which will be donated to the Tom Thorne and Beth Williams Memorial Award Fund (See Supplement to JWD, April, 2006) and the rest will go toward student support and activities. Later at the banquet, attendees were privileged to witness the bestowing of the Distinguished Service Award upon Dr. Paul Barrows and the Emeritus Award upon Dr. Robert Rausch (See below). In addition, one could say that the WDA continues to make definite strides in certain areas, such as with the coveted Duck Award, which was earned by Dr. Cynthia Tate, only the second woman ever to achieve such prestigious recognition in the Duck’s 17-year history. Many thanks to Dr. Richard French and the folks at UCONN for putting on such a fantastic meeting! We hope to see everyone again next year in Estes Park, Colorado, USA!

Dr. Paul L. Barrows receives 2006 Distinguished Service Award. Dr. Paul Barrows is this year’s Distinguished Service Award recipient. Paul has had a truly unique manner in which he has contributed a great deal to the WDA. The mission of the WDA is to “acquire, disseminate and apply knowledge” about health of wildlife. Many of us contribute mainly to the acquisition of knowledge and then dissemination through publication and talks about our research. Although Paul has conducted research and has published results of it, it is through his commitment to the dissemination and application of knowledge about the health and management of wildlife as influenced by military and professional scientific organizations that Paul has contributed so significantly.

His service to the WDA is an excellent example of Paul’s willingness to serve as a team member within professional wildlife organizations. Paul was the Chair of the ad hoc committee on mycoplasmosis in wild turkeys in 1986; member of the Promotions Committee, ’88–’91 [Chair 1989–1991]; member of the Public Awareness Committee [1993–1995]; and member and Chair of the LongTerm Vision Committee [2004–2005].

Paul was first on the WDA Council where he did an exemplary job as Secretary from 1990 to 1993. Paul was elected back to Council as Member-At-Large from 1997 to 2000. From July, 2001 to August, 2003, Paul was President of the WDA and he continued on Council as Past President through to August 2005.

Not only has Paul spent a great deal of time serving the WDA, he has been a leading part of a team of WDA members that has brought significant change to the Association. During the period that Paul was President, the WDA Council pursued the goal that led to the WDA publishing an electronic version
of the *Journal of Wildlife Diseases*. Under Paul's direction, the Council took the bold step to develop and fill the position of executive manager for the Association. This decision occurred approximately 10 years after the suggestion was introduced by David Jessup. In addition, the Council under Paul's direction took the very bold and successful decision to raise subscription and membership fees significantly to help defray the expenses of added initiatives. This action placed the WDA in its most favorable financial position of the previous 50+ years. At this time, Paul also put great effort into trying to interest corporate sponsorship in the WDA. Despite lackluster enthusiasm on the part of the corporations approached, Paul's efforts seemed boundless.

Paul will be the first person to remind us that these many significant achievements of which he was a part were indeed just that, an achievement of teams of many likeminded people working together. It is this clear commitment to the benefits of team work and his ability to work well as a team member that have led to Paul making major contributions in, not only the WDA, but also while serving on the governing councils and/or committees of other scientific societies such as the American College of Veterinary Preventive Medicine, American Association of Wildlife Veterinarians and the American Veterinary Medical Association.

Paul was a member of the United States Army for many years, retiring as a Colonel with the U.S. Army Veterinary Corps. In the latter stages of his army career, Paul was Commander of the U.S. Army Veterinary Command at Fort Sam Houston, Texas [1993–1996]; Chief of the Veterinary Corps [1995–1999]; Director of the Department of Defense, Veterinary Service Activity [1995–1999]; and Chief, Department of Veterinary Science, Army Medical Department Center and School, Fort Sam Houston, TX [1996–2000].

Paul's contributions for his service in his profession have been acknowledged by his university, scientific societies and the military. Paul was recognized in 1996 with the Distinguished Veterinary Alumnus Award of Michigan State University and in 2001 with the Michigan State University Distinguished Alumnus Award. Paul is a recipient of the Helwig-Jennings Award of the American College of Veterinary Preventive Medicine for significant and lasting contributions to the totality of veterinary preventive medicine.

Paul's contributions to dissemination and application of science about health of wildlife within the military have not only been reflected in the positions to which he was appointed within the U.S. Army, but also with a plethora of medals and other acknowledgements. Congratulations Dr. Paul Barrows!

-Submitted by Ed Addison

**Dr. Robert L. Rausch receives 2006 WDA Emeritus Award.** Dr. Bob Rausch, recipient of the 2006 WDA Emeritus Award, has been a role model to many young scientists working in the area of wildlife...
health and parasitology during the last half of the 20th century. Bob Rausch received his BA and DVM at Ohio State University and carried on to complete an MS at Michigan State before completing his PhD in parasitology and wildlife management at the University of Wisconsin in 1949.

After receiving his PhD, Bob began 25 years of a highly productive and illustrious career in research at the Arctic Health Research Institute of the US Public Health Service in Alaska. During his work in Alaska, Bob used his knowledge of Russian to collaborate on numerous research projects with scientists of the former Soviet Union. While this, of itself, was quite unique, it is an example of Bob’s career long commitment to work with others for the advancement of science. By the time that Bob had left his post in Alaska in 1975, he was an author of more than 170 research papers. While the focus of Bob’s research was wildlife helminthology with a special emphasis on the Cestoda, Bob displayed through his publications very diverse interests in natural history. These included papers on such topics as methods of handling wildlife, pathology associated with starvation in deer, dentition in ursids, genetics of mammals, distribution of fauna, and morphometry and reproduction in wolverines. While a highly productive helminthologist, Bob has certainly retained a broad interest in natural history!

After Alaska, Bob took a position at the University of Saskatchewan, a location he and his wife enjoyed immensely for a few years. However, he was forced to leave that position due to some administrative ‘red tape’ about him potentially forfeiting his U.S. government pension if he continued to work outside of his native country of subjects about which he acquired experience while in the U.S. public service. Before leaving Saskatoon, Bob and his wife Virginia established the Robert L. and Virginia R. Rausch Visiting Professorship at the Western College of Veterinary Medicine. This initiative has made a large contribution to promoting the exchange of information and experience within the wildlife health community.

From 1978 to 1992, Bob held a variety of positions at the University of Washington in Seattle, including Professor of Pathobiology and Professor of Comparative Medicine. Since 1992, Bob has been Professor Emeritus and continues to publish. By 2004, Bob had been an author of at least 280 papers.

With such a productive and uniquely varied career, it comes as no surprise that Bob has been recognized many times for his efforts. Amongst numerous other awards, Bob received the prestigious Henry Baldwin Ward Medal of the American Society of Parasitologists in 1961 and the Meritorious Service Medal of the U.S. Public Health Service in 1965. Bob has received a number of honorary degrees and he has received the Distinguished Service Award of both the American Society of Parasitologists [2001] and the Wildlife Disease Association [1983].

Bob has contributed very extensively to science organizations. To cite only a couple of activities, Bob is a charter member of the WDA. He was a member of the WDA Council [1976–1978] and filled many positions, including that of President [1984] of the American Society of Parasitologists. Bob also has contributed as a member to the editorial board of a number of journals.

Bob has always seen things from a broad perspective. He is an ecologist, naturalist, scholar, and
a gentleman. He has greatly influenced many of us who have chosen a similar career path and is very deserving of this recognition. **Congratulations to Dr. Bob Rausch!**

-Submitted by Ed Addison

**WDA STUDENT ACTIVITIES**

**STUDENT NEWS**

**European Student Chapter of the Wildlife Disease Association.** We are pleased to announce the creation of the **EWDA Student Chapter**, the European Student Chapter of the Wildlife Disease Association! With a prime objective to promote shared knowledge between established researchers and wildlife disease students. Subscribe to the **EWDA discussion E-list**, learn about the latest wildlife health and disease news, job and education opportunities, and discuss and debate wildlife health hot topics! Participate in the **EWDA Student Workshop**, enhance your wildlife disease research skills, and meet renowned scientists and potential mentors! Don’t miss the **EWDA Conference Student Mixer**, develop international relationships with students from all over Europe and beyond, and have fun!

Whether you are undergraduate, MSc, PhD, or post-doc, become TODAY a EWDA Student Chapter MEMBER! Membership form and all you need to know are on the student page of the EWDA website (European section of the Wildlife Disease Association: www.ewda.org).

- **Leslie A Reperant, WDA and EWDA Student Representative (reperant@Princeton.EDU).**

**STUDENT AWARDS**

**Storrs, Connecticut, USA 2006.** For this year’s student awards, Dr. Todd Cornish and his fellow Student Awards Committee members had an exceptionally difficult task in judging the Graduate Student Research Recognition Award and the Scholarship Award applicants. They were both extremely close races but two students managed to shine just a little brighter. As always, the Terry Amundson Student Presentation Competition also attracted many excellent students. Thanks to all the folks who helped with judging the session!

Congratulations to all the students who participated in this year’s student competitions!

1. **WDA Graduate Student Research Recognition Award**

**WINNER:** Justin Brown  
Southeastern Cooperative Wildlife Disease Study, Department of Population Health, College of Veterinary Medicine, University of Georgia  
“Persistence of H5 and H7 Avian Influenza Viruses in Water”  
Awarded plaque and $2,000 (US) for travel to WDA meeting

2. **WDA Scholarship Award**

**WINNER:** Krysten Schuler  
Department of Wildlife and Fisheries Sciences, South Dakota State University  
Awarded plaque and $2,000 (US) for educational expenses

3. **WDA Terry Amundson Student Presentation Award**

**WINNER:** Pauline Nol  
Department of Clinical Sciences, College of Veterinary Medicine and Biomedical Sciences, Colorado State University  
“Efficacy of Oral and Parenteral Bacille Calmette-Guerin (BCG Danish Strain 1331) in Protecting White-tailed Deer (Odocoileus virginianus) Against Bovine Tuberculosis”  
Awarded plaque and $250 (US)

**HONORABLE MENTION (3):**  
Claire Jardine  
Department of Veterinary Pathology, Western College of Veterinary Medicine, University of Saskatchewan
“Effect of Experimental Ectoparasite Control on Bartonella Infections in Wild Richardson’s Ground Squirrels (Spermophilus richardsoni)”
Awarded plaque and $75 (US)
Sonia Hernandez-Divers
Institute of Ecology, University of Georgia

“Investigating the Health and Pathogen Diversity of Backyard Chickens in Latin America”
Awarded plaque and $75 (US)
Christy Wyckoff
Caesar Kleberg Wildlife Research Institute, Texas A&M University – Kingsville

“Preliminary Evidence of Interactions Between Domestic Swine and Pseudorabies- and Brucellosis-Positive Feral Swine”
Awarded plaque and $75 (US)

ATTENTION MENTORS AND ADVISORS! Please encourage your students to apply for WDA’s student awards. Each year the WDA sponsors student awards competitions. For 2007, students are encouraged to apply for three awards. The WDA Student Awards Committee (comprised of 8 members from around the globe) will judge the Graduate Student Research Recognition Award and Scholarship Award. Members of the audience attending the annual WDA meeting will judge the Terry Amundson Student Presentation Award. Criteria for judging of the awards are available on the WDA website at the following URL: http://www.wildlifedisease.org/Student_Awards.htm.

Applicants MUST be Student Members of the WDA at the time applications for the Graduate Student Research Recognition and Scholarship Awards are received and/or at the time the abstract for the Terry Amundson Student Presentation Award is submitted for consideration. Furthermore, students applying for the Graduate Student Research Recognition Award and Scholarship Award MUST be pursuing an advanced degree at the time of application.
## Happenings in the Field

USGS National Wildlife Heath Center’s Quarterly Wildlife Mortality Report

### Quarterly Wildlife Mortality Report

January 2006 to March 2006

<table>
<thead>
<tr>
<th>State</th>
<th>Location</th>
<th>Dates</th>
<th>Species</th>
<th>Mortality</th>
<th>Diagnosis</th>
<th>Reported by</th>
</tr>
</thead>
<tbody>
<tr>
<td>GA</td>
<td>Miller County</td>
<td>03/21/06–ongoing</td>
<td>River Frog</td>
<td>50 (e)</td>
<td>Parasitism: Perkinsus-like organism</td>
<td>NW SCW</td>
</tr>
<tr>
<td>AR</td>
<td>Bald Knob NWR, North Carolina</td>
<td>01/06/06–01/15/06</td>
<td>Greater Snow Goose, Ross’s Goose</td>
<td>1,000 (e)</td>
<td>Aflatoxicosis suspect</td>
<td>NW</td>
</tr>
<tr>
<td>BC</td>
<td>East Coast of Queen Charlotte Islands</td>
<td>12/25/05–04/15/06</td>
<td>Cassin’s Auklet</td>
<td>100 (e)</td>
<td>Emaciation: starvation</td>
<td>BC WT</td>
</tr>
<tr>
<td>CA</td>
<td>Silver Strand State Beach</td>
<td>03/15/06–03/16/06</td>
<td>Western Grebe</td>
<td>70</td>
<td>Trauma</td>
<td>SDC, SWD</td>
</tr>
<tr>
<td>CA</td>
<td>Tule Lake NWR and Lower Klamath NWR</td>
<td>03/06/06–04/10/06</td>
<td>Ring-Necked Duck, Bufflehead, Ruddy Duck, Snow (Blue-phase) Goose, American Coot, Unidentified Gull</td>
<td>600 (e)</td>
<td>Undetermined Avian Cholera</td>
<td>NW</td>
</tr>
<tr>
<td>FL</td>
<td>Daytona Beach</td>
<td>12/27/05–01/20/06</td>
<td>Unidentified Gull, Herring Gull, Laughing Gull, Ring-Billed Gull, Mallard</td>
<td>25 (e)</td>
<td>Aspergillosis</td>
<td>KDL SCW</td>
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<tr>
<td>FL</td>
<td>Fountain</td>
<td>01/24/06–02/25/06</td>
<td>Green Frog</td>
<td>150 (e)</td>
<td>Viral Infection (suspect): Iridovirus</td>
<td>NW</td>
</tr>
<tr>
<td>GA</td>
<td>Hoginsville</td>
<td>03/25/06–03/25/06</td>
<td>Black Vulture, Dog, Eastern Cottontail, Red-Winged Blackbird</td>
<td>4</td>
<td>Toxocosis: aldicarb</td>
<td>SCW</td>
</tr>
<tr>
<td>GA</td>
<td>Macon</td>
<td>12/30/05–12/30/05</td>
<td>Lesser Snow Goose, American Coot, Canada Goose, Trumpeter Swan, Long-Tailed Duck</td>
<td>7</td>
<td>Undetermined Esophageal ulcers, Hepatitis</td>
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<tr>
<td>ID</td>
<td>Camas NWR</td>
<td>03/26/06–04/10/06</td>
<td>Ross’s Goose, Lesser Snow Goose, American Coot, Canada Goose, Tundra Swan, Greater Sandhill Crane</td>
<td>437</td>
<td>Aflatoxicosis suspect</td>
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<tr>
<td>IL</td>
<td>Wilmette</td>
<td>02/15/06–03/02/06</td>
<td>Greater Sandhill Crane</td>
<td>5 (e)</td>
<td>Open</td>
<td>NW</td>
</tr>
<tr>
<td>KS</td>
<td>Quivira NWR</td>
<td>03/08/06–03/11/06</td>
<td>Greater Sandhill Crane</td>
<td>200 (e)</td>
<td>Mycotoxicosis suspect</td>
<td>NW</td>
</tr>
<tr>
<td>MN</td>
<td>Lake Winnibigoshish</td>
<td>10/07/05–10/13/06</td>
<td>American Coot, Greater Sandhill Crane</td>
<td>40 (e)</td>
<td>Open</td>
<td>NW</td>
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<tr>
<td>MN</td>
<td>Mississippi River, above Lock 4</td>
<td>10/10/05–10/15/05</td>
<td>American Coot, Greater Sandhill Crane</td>
<td>200 (e)</td>
<td>Trauma: water intake pipes</td>
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<tr>
<td>MO</td>
<td>Bob Brown State Conservation Area</td>
<td>01/15/06–01/31/06</td>
<td>Greater Snow Goose</td>
<td>183</td>
<td>Avian cholera</td>
<td>NW</td>
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<tr>
<td>MO</td>
<td>Squaw Creek NWR</td>
<td>01/15/06–01/31/06</td>
<td>Greater Snow Goose, Mallard, Ross’s Goose</td>
<td>150</td>
<td>Avian cholera</td>
<td>NW</td>
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<tr>
<td>OR</td>
<td>OR Coast, Columbia River to Port Orford</td>
<td>03/06/06–04/07/06</td>
<td>Rhinoceros Auklet</td>
<td>400 (e)</td>
<td>Emaciation</td>
<td>NW</td>
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<td>OR</td>
<td>Sauvie Island Wildlife Area</td>
<td>01/01/06–03/15/06</td>
<td>Lesser Snow Goose, Canada-Dusky Goose, Cackling Goose</td>
<td>37</td>
<td>Lead Poisoning</td>
<td>NW</td>
</tr>
</tbody>
</table>
### AIV isolations from Wildbirds: Europe: Sept.2006–June.2006

The results of AIV surveillance in wild birds for European countries have been summarised by several organisations e.g. ProMED and in the web report below:


A summary for events in March 2006 is given below — Acknowledgements: As on source and and Thijs Kuiken. Please note that these data were produced in March 2006.

<table>
<thead>
<tr>
<th>State</th>
<th>Location</th>
<th>Dates</th>
<th>Species</th>
<th>Mortality</th>
<th>Diagnosis</th>
<th>Reported by</th>
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</thead>
<tbody>
<tr>
<td>OR</td>
<td>Briedwell Road, Amity</td>
<td>01/13/06–01/18/06</td>
<td>Cackling Goose</td>
<td>6</td>
<td>Toxicosis: CHE Inhibiting cmpd.</td>
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<tr>
<td>TX</td>
<td>Dalhart</td>
<td>01/06/06–01/17/06</td>
<td>Ross’ Goose</td>
<td>34</td>
<td>Open</td>
<td>NW</td>
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<tr>
<td>VA</td>
<td>2126 Lake View Dr., Powhatan</td>
<td>03/01/06–03/01/06</td>
<td>Cedar Waxwing Yellow-Bellied Sapsucker</td>
<td>4 (e)</td>
<td>Open: toxicosis suspect</td>
<td>SCW</td>
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<tr>
<td>WA</td>
<td>Coastal areas</td>
<td>12/01/05–04/30/06</td>
<td>Trumpeter Swan</td>
<td>400 (e)</td>
<td>Lead poisoning</td>
<td>BC, WA</td>
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<tr>
<td>WA</td>
<td>Seattle</td>
<td>03/18/06–03/18/06</td>
<td>Cedar Waxwing</td>
<td>18</td>
<td>Toxocosis: cyanide</td>
<td>NW</td>
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<tr>
<td>WA</td>
<td>Snake River, Clarkson</td>
<td>03/20/06–03/21/06</td>
<td>Mallard</td>
<td>8</td>
<td>Toxicosis: Famphur</td>
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<td>WI</td>
<td>Lake Onalaska, Pool 7, Upper Mississippi</td>
<td>03/20/06–ongoing</td>
<td>Lesser Scaup American Coot Ring-Necked Duck Canada Goose Greater Snow Goose</td>
<td>2,400 (e)</td>
<td>Parasitism: Sphaeridiotrema globalis</td>
<td>NW</td>
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**Updates:**

<table>
<thead>
<tr>
<th>State</th>
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<th>Diagnosis</th>
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<tr>
<td>NC</td>
<td>Moore Co., Woodlake</td>
<td>11/20/05–12/18/06</td>
<td>American Coot Bald Eagle Lesser Sandhill Crane</td>
<td>300 (e)</td>
<td>Vacular myelopathynthesis</td>
<td>NW, SCW</td>
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<tr>
<td>TX</td>
<td>Lorenzo</td>
<td>12/14/05–12/15/05</td>
<td>Lesser Sandhill Crane Ruddy Duck</td>
<td>34</td>
<td>Open: aflatoxicosis suspect</td>
<td>NW</td>
</tr>
<tr>
<td>TX</td>
<td>Muleshoe NWR</td>
<td>12/15/05–02/23/06</td>
<td>Lesser Sandhill Crane Ruddy Duck</td>
<td>400 (e)</td>
<td>Mycotoxicosis Trichothecene suspect</td>
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<tr>
<td>WI</td>
<td>Lake Onalaska, Upper Miss NWR</td>
<td>08/30/05–11/25/05</td>
<td>American Coot Lesser Scaup Blue-Winged Teal Ring-Necked Duck Mallard</td>
<td>8,850 (e)</td>
<td>Parasitism: Cyathocotyle bushiensis</td>
<td>NW</td>
</tr>
</tbody>
</table>

(e) = estimate; * = morbidity, not mortality

B.C. Ministry of Environment (BC), USGS National Wildlife Health Center (NW), Southeastern Cooperative Wildlife Disease Study (SCW), San Diego County Veterinary Diagnostic Laboratory (SDC), Sea World (SWD), Washington Animal Disease Diagnostic Laboratory (KDL), Wildlife Trust (WT). Written and compiled by Kathryn Converse/Rex Sohn - Western US, Grace McLaughlin - Eastern US, NWHC. The Quarterly Wildlife Mortality Report is available at http://www.nwhc.usgs.gov. To report mortality or receive information about this report, contact the above NWHC staff, or for Hawaiian Islands contact Thierry Work. Phone: (608) 270-2400, FAX: (608) 270-2415 or e-mail: kathy_converse@usgs.gov. USGS National Wildlife Health Center, 6006 Schroeder Road, Madison, WI 53711.

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**WDA SECTION NEWS**

**NEWS FROM THE EUROPEAN SECTION**

**EWDA WEBSITE - www.ewda.org** Visit the EWDA website at www.ewda.org and find information on conferences, members interests, publications and lots more. The website is kindly provided free of charge by the UK Central Science Laboratory. We are considering a new look for the site so if you have any further suggestions or material then please send them to r.delahay@csl.gov.uk.

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**AIV isolations from Wild birds: Europe: Sept.2006–June.2006** The results of AIV surveillance in wild birds for European countries have been summarised by several organisations e.g. ProMED and in the web report below:


A summary for events in March 2006 is given below — Acknowledgements: As on source and and Thijs Kuiken. Please note that these data were produced in March 2006.
Collaborators from Western Europe were asked for summaries of AIV surveillance in wild birds from their respective countries, and the responses, which we stress are not official data, are given below.

<table>
<thead>
<tr>
<th>Country</th>
<th>HPAI/H5N1 isolated?/region/isolates in which months</th>
<th>Species</th>
<th>HPAI – how many (geographical) foci of infection in your country. Regions affected</th>
<th>HPAI – date of last isolation</th>
<th>LPAI – how many isolates</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Austria</td>
<td>See summary below</td>
<td>N/a</td>
<td>N/a</td>
<td>N/a</td>
<td>1 × H3N6 (in a mallard)</td>
<td></td>
</tr>
<tr>
<td>Belgium</td>
<td>No</td>
<td>N/a</td>
<td>N/a</td>
<td>N/a</td>
<td>405 PCR positive samples</td>
<td>Period 1 July 2005 to 31 May 2006</td>
</tr>
<tr>
<td>Denmark</td>
<td>See summary</td>
<td>N/a</td>
<td>N/a</td>
<td>N/a</td>
<td></td>
<td></td>
</tr>
<tr>
<td>France</td>
<td>See summary</td>
<td>N/a</td>
<td>N/a</td>
<td>N/a</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Germany</td>
<td>See summary</td>
<td>N/a</td>
<td>N/a</td>
<td>N/a</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Great Britain</td>
<td>H5N1 ×1 Scotland April</td>
<td>Whooper swan</td>
<td>1 focus Scotland</td>
<td>April 2006</td>
<td>6 isolates (see <a href="http://www.Defra.uk">www.Defra.uk</a>)</td>
<td>Origin of H5 not known</td>
</tr>
<tr>
<td>Netherlands</td>
<td>No</td>
<td>N/a</td>
<td>N/a</td>
<td>N/a</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Italy</td>
<td>H5N1 Puglia, Calabria,Sicilia, Umbria February</td>
<td>Mute swan;Purple swamphen; Buzzard; Mallard</td>
<td>6×Puglia, 2×Calabria,7×Sicilia, 1×Umbria</td>
<td>February 2006</td>
<td>7 isolates</td>
<td>Origin of H5 not known</td>
</tr>
<tr>
<td>Spain</td>
<td>H5N1 ×1 Saburua lake, Alava</td>
<td>Great crested grebe</td>
<td>Alava × 1</td>
<td>June 2006</td>
<td>N/a</td>
<td></td>
</tr>
<tr>
<td>Sweden</td>
<td>H5N1, along east coast of Sweden, from southern tip up to, and including Stockholm region, from Feb to April</td>
<td>Tufted duck, Mute swan, Goosander, Scaup, Canada goose, Smew, Herring gull, Mallard, Common buzzard, European eagle owl, American Mink</td>
<td>12</td>
<td>21 April 2006</td>
<td>18 PCR positive samples from 250 healthy farmed Mallards</td>
<td></td>
</tr>
</tbody>
</table>

H5N1 virus surveillance in wild birds
EU winter 05–06

-22 member states
-534 positives in 14 countries
-24,093 birds tested
-1060 Influenza A virus positive
-No HPAI virus found

### Austria:
So far we have examined more than 3,295 wild birds that have been found dead and because of legal provisions been sent to the competent authority. We have detected H5N1 in 125 wild birds.

The table shows the species affected against the calendar weeks:

<table>
<thead>
<tr>
<th>Calendar week</th>
<th>Coot</th>
<th>Wild duck</th>
<th>Wild goose</th>
<th>Chicken</th>
<th>Gull</th>
<th>Heron</th>
<th>Swan</th>
<th>Waterfowl spp.</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>7</td>
<td></td>
<td>12</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>6</td>
<td>8</td>
<td>2</td>
<td>28</td>
</tr>
<tr>
<td>8</td>
<td></td>
<td>2</td>
<td>4</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>9</td>
<td></td>
<td>2</td>
<td>3</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>5</td>
<td>1</td>
<td>12</td>
</tr>
<tr>
<td>10</td>
<td></td>
<td>3</td>
<td>3</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>10</td>
<td>25</td>
<td>534</td>
</tr>
<tr>
<td>11</td>
<td></td>
<td>1</td>
<td>2</td>
<td>1</td>
<td>6</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>10</td>
</tr>
<tr>
<td>12</td>
<td></td>
<td>1</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>12</td>
</tr>
<tr>
<td>13</td>
<td></td>
<td>2</td>
<td>5</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>10</td>
<td>7</td>
<td>3</td>
</tr>
<tr>
<td>14</td>
<td></td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>1</td>
<td>5</td>
</tr>
<tr>
<td>15</td>
<td></td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>16</td>
<td></td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>17</td>
<td></td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>10</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>29</td>
<td>3</td>
<td>7</td>
<td>7</td>
<td>5</td>
<td>12</td>
<td>16</td>
<td>25</td>
</tr>
</tbody>
</table>

The results of the screening program in wild birds can however not be published yet.

### Germany:
Since February 15th, a total of 343 wild birds, three stray cats and one stone marten have been identified in Germany as being infected with the HPAIV H5N1 of the so-called Qinghai lineage. All cases were identified through passive monitoring, i.e. by examining more than 40,000 dead birds/animals. Since the second week of May no new cases have been reported in wild birds, however, passive monitoring intensities are regressing constantly since early May. A single outbreak of HPAIV H5N1 has occured in a mixed poultry holding in early April in the federal state of Saxonia demanding the cul-
ling of approximately 25,000 birds in the restriction zone. No further outbreaks in poultry have been noticed since. The poultry isolate showed a very close relationship to H5N1 viruses circulating at that time in the wild bird population of north-eastern Germany.

**Netherlands:** Between 1 July 2005 and 31 May 2006, the Erasmus MC in Rotterdam examined swabs from 10,086 apparently healthy wild birds by RT-PCR, of which 405 were positive for low pathogenic influenza A virus and none for high pathogenic influenza A virus. Up to 24th April 2006, the Central Institute for Animal Disease Control in Lelystad examined 6,288 wild bird carcasses, none of which were positive for high pathogenic influenza A virus.

**Spain:** In Spain active and passive surveillance for HPAI is in place for poultry and wild birds. Sample collection and submission is handled differently in different communities. In most communities the sampling effort is above the numbers set by the ministry of agriculture. All samples are analysed by the central veterinary laboratory however in some communities parallel testing is in place. Sampling of wild birds has declined after the spring migration. A Great Crested Grebe (*Podiceps cristatus*) found dead at the Salburua lake near Vitoria in the province of Alava (Northern Spain) has tested positive for H5N1. Apparently the strain shows similarities to others isolated in Northern Europe. The case has alerted the public and caused passive surveillance to pick up again with more carcasses reported by the public.

**France:** In France, since February 13th, HP H5N1 of the so-called Qinghai lineage was isolated from 42 pools corresponding to 66 dead birds. 83% of the infected birds were mute swans (*Cygnus olor*). 41 pools came from the “département” of Ain (most of them from the famous wetland of Dombes, located in the north-east of the city of Lyon):

- 32 positive pools of Mute Swans corresponding to 32 to 54 dead birds (some birds were tested alone, some others by pools of 2, 3, 4 or 5 birds)
- 4 positive pools of Common Pochards (*Aythya ferina*), corresponding to 6 dead birds
- 1 positive Tufted duck (*Aythya fuligula*)
- 1 positive Common Buzzard (*Buteo buteo*)
- 1 positive Grey Heron (*Ardea cinerea*)
- 1 positive Greylag goose (*Anser anser*)
- 1 positive Great Crested Grebe (*Podiceps cristatus*)

Only one dead Mute Swan was found positive, the 28th February, out of the Ain, in the “département” of the Bouches du Rhône, in the South of France, near the city of Marseille.

All cases were identified in February, March and April, through passive monitoring, i.e. by testing more than 3,000 dead birds. Since the 18th of April no new cases have been reported in wild birds in the Dombes. A single outbreak of HPAIV H5N1 has occurred in a turkey farm (the 23th February), very close to the first cases identified in Common Pochard on the 13th February. No further outbreaks in poultry have been recorded since that time.

Concerning the active survey performed since September 2005 throughout France, more than 4,000 captured or hunter-killed birds were analysed (*Anatidae, Laridae, corvidae*) from which no HP H5N1 was isolated. Approximately one hundred isolates of LPAI strains from wild birds were identified, of which 9 were H5 LP non-N1, and one was H7N1 LP.

**Sweden:** Passive surveillance of dead wild birds resulted in finding the HPAI positive birds in Sweden. Of 519 PCR-tested bird carcasses, 68 tracheal swabs were found positive for H5 avian influenza. The birds were all water-living species, or predators feeding on these birds. There were no mass die-offs, only single or very low numbers of dead birds were found within the surveillance zones, in spite of large congregations of birds. A few HPAI positive birds were seen showing neurologic signs, mainly as uncontrolled “spinning”, i.e. swimming in tight circles on the water. One wild mink with neurological signs found within one of the surveillance zones, was euthanased, and was found positive for H5 avian influenza on PCR. The positive cases were found along the ice-free areas of the eastern coast where birds congregated whilst waiting for the spring thaw. As soon as warmer weather arrived in late April, the birds dispersed to their breeding grounds, and no further positive cases were found. One game farm with mallards and pheasants within a surveillance zone was tested, and from the samples a single healthy mallard was positive for HPAI, which led to destruction of all birds at that farm. LPAI was found at this farm as well. There were no outbreaks of avian influenza in Swedish poultry farms. Active surveillance of migratory birds is continuing, but has so far not shown any finding of HPAI.

**Denmark:** The first case of HP H5N1 in Denmark was a common buzzard (*Buteo buteo*) found dead in the southern part of Sealand 13 March 2006. Since then the Danish Institute for Food and Veterinary Research has examined samples from approximately 1,300 wild birds and HP H5N1 has been detected.
in samples from a total of 44 birds. The positive birds have primarily been Tufted ducks (Aythya fuligula), swans and raptors (mainly buzzards). Positive findings have mainly been located in the Baltic region, Roskilde fjord and the northern part of Sealand. All cases in wild birds were identified through passive monitoring, i.e. birds found sick or dead and submitted for analysis. Until now there has been only one outbreak of HP H5N1 in domestic poultry. This outbreak was identified 18th May and involved a mixed poultry holding on the Island of Funen. No further outbreaks in poultry have been identified since. The last positive finding in Danish wild birds was a magpie (Pica pica) found dead on the property of the mixed poultry holding with the HP H5N1 outbreak 23 May. More information concerning HP H5N1 analyses of Danish wild birds is available from the official website of the Danish Institute for Food and Veterinary Research:
http://www.dfvf.dk/Files/Filer/Husdyrsygdomme/Avi%20influenza/AI_resultater.xls

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Australasian Section. For information regarding the Australasian Section, contact Pam Whiteley, 1 Brinsley Road, Camberwell, VIC 3124, Australia. Telephone: 61-3-98825608; Fax: 61-3-98823054; E-mail: pwhitele@bigpond.net.au.

European Section. For information regarding the European Section, contact Dolores Gavier-Widen, Division of Wildlife Fish and Environment, SVA, SE 751 89 Uppsala, Sweden, Telephone: 46-18-674-215; Fax: 46-18-30-91-62; E-mail: dolores@sva.se.

Nordic Section. For information regarding the Nordic Section, contact Eric Agren, Department of Wildlife, National Veterinary Institute, SE-751 89 Uppsala, SWEDEN, Telephone +46 18 67 40 00 Fax +46 18 30 91 62 or E-mail: Erik.Agren@sva.se.

Wildlife Veterinarian Section. For information regarding the Wildlife Veterinarian Section, contact Kirsten Gilardi, Wildlife Health Center, University of California Davis, CA 95616 USA. Telephone: 530-752-4896, FAX: 530-752-3318, email: kvgilardi@ucdavis.edu.

JOB ANNOUNCEMENTS

Visit the JWD website at http://www.wildlifedisease.org/Jobs.htm for up to date job listings.

TRAINING/EDUCATIONAL OPPORTUNITIES

Visit the JWD website at http://www.wildlifedisease.org/Training.htm for more information on training opportunities.

Avian, Reptile, Rabbit, Ferret, and Rodent Diagnostic Endoscopy Course. November 18 and 19, 2006. This 15 hour continuing education course is designed to teach the theory and practical applications of diagnostic endoscopy in birds, reptiles and small mammals. Whether you are a private practitioner, zoo/aquarium/wildlife veterinarian, or researcher this course will train you to perform minimally-invasive endoscopic procedures including biopsy techniques. This is a basic to intermediate level course, and fundamental knowledge of avian and reptilian anatomy is assumed. You will be trained using PowerPoint lectures and video presentations in equipment choice and care; oral, aural, and upper respiratory endoscopy in small mammals; colendoscopy, gastro-intestinal and respiratory endoscopy of reptiles; colendoscopy, gastro-intestinal endoscopy and tracheoscopy of birds; biopsy and sampling techniques; and endoscopy fee structure and practice management. In addition, there will be over 8 hours of practical lab time in which you will be able to practice and develop your skills in anesthetized (non-recovery) research iguanas and pigeons scheduled for euthanasia. All procedures approved by the UGA Institutional Animal Care and Use Committee. Refreshments, lunches, certificate of training, and full color printed course notes
Fish and Chelonian Diagnostic Endoscopy Course. December 2 and 3, 2006. This 15 hour continuing education course is designed to teach the theory and practical applications of diagnostic endoscopy in fish (including koi and catfish) and chelonians (including turtles, tortoises and terrapins). Whether you are a private practitioner, zoo/aquarium/wildlife veterinarian, or researcher this course will train you to perform minimally-invasive endoscopic procedures including biopsy techniques in fish and turtles. The course will include PowerPoint video lectures and practical instruction in gastro-intestinal, respiratory, and coelomic endoscopy. Special emphasis will be placed upon equipment selection, animal preparation, endoscopic identification of tissues and organs, and the collection of biological samples for disease diagnosis and research purposes. In addition, there will be over 8 hours of practical wet-lab time to practice and develop endoscopy skills in anesthetized (non-recovery) farmed koi, catfish and red-eared sliders (terrapins). All procedures approved by the UGA Institutional Animal Care and Use Committee. Refreshments, lunch, certificate of training, and full color printed course notes containing all tutorial materials will be provided. Limited to 16 veterinarians. $750 for veterinarians, $150 for accompanying technicians. Course will be held in surgery suites at the College of Veterinary Medicine, University of Georgia, Athens, GA 30602, USA. The college is 1.5 hrs shuttle bus ride from the Atlanta International Airport, and only 10 minutes from the Athens Regional Airport with service from the Charlotte International Airport. For registration details contact Sandi Kilgo at Telephone: 1-706-542-1451 or Email: skilgo@vet.uga.edu. For more information, visit the following website: www.gactr.uga.edu/conferences.

Dallas Zoo and Dallas Aquarium Veterinary Student Preceptorship A four to eight-week preceptorship offers exposure to clinical zoo and aquarium veterinary practice at a large metropolitan zoo. The student will work closely with the veterinary and keeper staff and receive an introduction to husbandry, restraint/immobilization, basic medical procedure techniques, and necropsies of zoo animals, the unique aspects of veterinary management of animals in a zoo setting, and the MedARKS recordkeeping system. An onsite library is available for use. Responsibilities will be assigned based on the student’s areas of interest and experience level. The student is expected to complete a project and present results to the veterinary staff, and will be responsible for local transportation, housing, and food. Applicants should be a fourth year veterinary student (or in final year of non-U.S. veterinary program) and have completed four weeks of a clinical medicine or surgery rotation before the start of the preceptorship. Negative tuberculin skin test within 60 days of the start of the preceptorship, current tetanus vaccination, and personal health insurance are required. Applicants should send a letter of intent, curriculum vitae, contact information for three references, and the name of their Zoo/Exotic Animal advisor to: Tim Storms, Associate Veterinarian at Dallas Zoo and Dallas Aquarium, 650 South R.L. Thornton Fwy., Dallas, Texas 75203-2996.

Training Available in Fish Diagnostics, Inspections, and Laboratory Methods. The US Fish and Wildlife Service Fish Health Centers provide laboratory and field examination services to the National Fish Hatcheries. Our main emphasis is to assist the hatcheries in producing quality fish that will contribute to the enhancement and restoration of aquatic ecosystems. At the Olympia and Idaho Fish Health Centers, the work may involve travel to field sites to perform diagnostic examinations and collect samples that are then evaluated in our laboratories. Routine testing procedures include bacteriology (biochemical, ELISA, and PCR methods), virology (cell culture, serological, and PCR methods), parasitology (microscopic and PCR methods), histology, and clinical chemistry. Training may be arranged for one day or several weeks at one or both of these laboratories depending on the interests and availability of the individual. In general, most broodstock inspections are performed from September through November, juvenile inspections are performed from January through April, and wild fish surveys are conducted from March through September. Routine diagnostic examinations are performed year round and special projects are conducted as time and necessity permit. For more information, please contact Joy Evered DVM, at the Olympia Fish Health Center; email joy.evered@fws.gov or Marilyn Blair DVM, at the Idaho Fish Health Center; email marilyn.j.blair@fws.gov.

Sr. Veterinary Student Preceptorship in Avian and Conservation Medicine. A four to six-week preceptorship in Avian and Conservation Medicine is being offered to a senior-year veterinary student by the International Crane Foundation (ICF) in Baraboo, Wisconsin. The preceptor will train with the Veterinary Services Unit of the Conservation Services Department in all phases of the clinical practice, but have opportunities for interaction with the Crane Conservation Department to learn captive propagation, hus-
bandry, and management of this unique family of birds. The preceptor can expect to gain practical experience in crane capture, transport, anesthesia, preventive medicine, disease surveillance, and the contribution of veterinary medicine to crane conservation including field project support and professional consultations. Preceptors are encouraged to complete and report on a research or laboratory project during their stay. Opportunities for visiting the University of Wisconsin School of Veterinary Medicine and the National Wildlife Health Center in Madison, WI will be made available to interested preceptors. No stipend is available for this position; however, on-site housing in the ICF Guesthouse will be provided depending on availability at the time the preceptorship is scheduled. Applicants should send a cover letter, curriculum vitae, or resume and one letter of recommendation from a faculty member of their home institution to: Barry Hartup, DVM, Director of Veterinary Services, International Crane Foundation, E-11376 Shady Lane Road, Baraboo, WI 53913, email hartup@savingcranes.org. Please view our website at www.savingcranes.org.

Directory of Post-Graduate Educational Opportunities in Zoo and Wildlife Medicine. The World Association of Wildlife Veterinarians has recently produced a Directory of Post-Graduate Educational Opportunities in Zoo and Wildlife Medicine. The Directory covers opportunities in over fifty countries and is a must for veterinary students or graduates interested in furthering their careers in the field of wildlife medicine. For further information, please contact the Secretary of the WAWV at: F.Scullion@zoo.co.uk.

MEETING AND CONFERENCE ANNOUNCEMENTS

Visit the JWD website at http://www.wildlifedisease.org for more conference listings.

Biological Crisis Management (BCM) in Human and Veterinary Medicines 2006. Emerging Diseases: Preparedness and Implementation Issues. Lyon, France. 5–8 Nov 2006. The Biological Crisis Management in Human and Veterinary Medicines 2006 Conference will be held in Lyon, France, November 5–8, 2006. The meeting is being organised by the International Association for Biologicals (IABs) in association with WHO, NIAID and OIE. As the first meeting of a new series on Biological Crisis Management in Human and Veterinary Medicines, BCM 2006 will bring together experts and representatives from industry and regulatory bodies to discuss important public health problems related to emerging infectious diseases. The goal of BCM 2006 is to help assure the rapid availability of safe and effective biologicals for use in emergency situations by identifying safety, efficacy, standardisation, liability and regulatory issues related to their production and use and to propose solutions. The meeting aims to provide recommendations for the scientific community, manufacturers and health authorities concerning the production, evaluation, standardisation and regulation of biologicals for use in emergency situations. For comprehensive information about the meeting (registration, accommodation, fees, programme, etc.), please visit the BCM2006 website: http://www bcm2006.org. For further information contact: Michele Michaud Dodet Bioscience, Conference Organiser, France; tel: +33 (0)4 72 41 17 06; fax: +33 (0)4 72 41 17 14; 66 cours Charlemagne 69002 Lyon, France; tel: (33) 4 72 41 17 08; email: BCM@dodetbioscience.com.

8th International Meeting Molecular Epidemiology and Evolutionary Genetics of Infectious Diseases (MEEGID VIII), Bangkok Thailand, November 30–December 4, 2006. The 8th International Meeting Molecular Epidemiology and Evolutionary Genetics of Infectious Diseases (MEEGID VIII) will be held in Bangkok, Thailand, 30 Nov–4 Dec 2006. As for the 7 first MEEGID meetings, it will be co-organized by the Centers for Disease Control and Prevention (http://www.edc.gov/) in Atlanta, the Centre National de la Recherche Scientifique (http://www.cnrs.fr/) and the Institut de Recherche pour le Développement (http://www.ird.fr/) in France, Mahidol University (http://www.mahidol.ac.th/) will be an official co-organizer of the meeting, which will be supported also by the French Embassy (http://www.ambafrance-th.org/). The MEEGID meetings are organized in synergy with the new journal Infection, Genetics and Evolution (Elsevier; http://www.elsevier.com/locate/meegid). Communications on genetics, genomics, proteomics, population biology, mathematical modelling, bioinformatics are welcome. They can deal with the host, the pathogen or the vector. Papers considering host/pathogen or pathogen/vector (co-evolution) are particularly encouraged. All pathogens are within the scope of MEEGID: viruses, parasitic protozoa, helminths, fungal organisms, prions. All infectious models can be considered, including those of veterinary or agronomical relevance. The papers accepted for MEEGID VIII will be published in a special issue of Infection, Genetics and Evolution, as already done for MEEGID VI (Paris, July 2002). Special emphasis will be given to health problems of special interest to Thailand and South-East Asia: avian flu, SARS, malaria, dengue. Contact: Michel Tibayrenc, MD, PhD, Editor-in-chief Infection, Genetics and Evolution (Elsevier), IRD representative in Thailand, IRD Representative Office, French Embassy, 29, Thanon Sathorn Tai, Bangkok 10120, Thailand, email Michel.Tibayrenc@ird.fr, (secretary) ird.th@ksc.th.com, Website: http://www.th.ird.fr, http://www.elsevier.com/locate/meegid.
The International Meeting on Emerging Diseases and Surveillance (IMED 2007), will be held in Vienna, Austria, February 23–25, 2007. This meeting is being co-sponsored by ProMED, the European CDC, the OIE (World Organisation for Animal Health), the WHO Regional Office for Europe, and the European Commission. IMED 2007 will bring together leaders in human and veterinary medicine and public health, epidemiology and surveillance, emergency preparedness, microbiology and other related disciplines to help improve our abilities to detect, monitor, and respond to emerging disease threats. The meeting will embrace the One Medicine concept, recognizing that, just as diseases reach across national boundaries, so do they cross species barriers. We therefore welcome the full participation of both the human and animal health communities. 2007, the year that the revised International Health Regulations will be fully implemented, is a highly fitting time to address the implementation of these broadly reaching regulations and to understand their implications for the surveillance of emerging diseases. Centrally located in Europe, the beautiful host city of Vienna is rich in history and culture. IMED 2007 will be organized by the International Society for Infectious Diseases, which has over 20 years experience in planning and implementing international biomedical meetings. With outstanding plenary speakers, symposia by expert panels, and invited abstracts, IMED 2007 is certain to be the year’s major conference for those involved in the study, detection, and monitoring of emerging pathogens and to those in the front lines of response. Please visit the IMED 2007 website http://imed.isid.org for more details and to take advantage of discounted early registration. Abstracts are invited and online abstract submission is available now.


Note from the Editor
Send any items such as reports, meeting announcements, diagnostic riddles, position and grant announcements, or anything else deemed appropriate for the Supplement to the Journal of Wildlife Diseases or the WDA website, to Pauline Nol (Supplement Editor) at USDA/APHIS, National Wildlife Research Center, 4101 LaPorte Avenue, Fort Collins, CO 80521 USA, Ph: (970) 266-6126, Fax: (970) 266-6157, Email: pauline.nol@aphis.usda.gov, or Michael Ziccardi (Website Editor) at Wildlife Health Center, University of California, Davis, CA, 95616, USA, Ph: (530) 754-5701, Fax: (530) 752-3318, Email: mhziccardi@ucdavis.edu. Files in Microsoft Word sent electronically or via disk are preferred, though submissions in any form are welcome. MANY THANKS!

—Pauline